



Effect of poultry production on air quality and human health in selected agricultural zones of Imo State



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ABSTRACT

The Confined Animal Feeding Operations (CAFOs) in poultry management are increasingly recognized as sources of air pollution, yet their health impacts remain underexplored. This study evaluated air pollutant levels and respiratory health among poultry workers in selected zones of the Poultry Association of Nigeria, Imo State, from December 2021 to May 2022. Air samples were collected monthly from six poultry pens across three zones. Pollutants measured included CO₂, CH₄, NO₂, NH₃, H₂S, SO₂, and PM_{2.5}, alongside microclimatic parameters such as temperature, humidity, and wind speed. Gaseous pollutants were assessed using Kanomax and iTX multigas analyzers, and PM_{2.5} levels were measured with a pDR-1200 Monitor. A structured questionnaire and spirometry tests were used to evaluate workers' health. Results showed mean NH₃ concentrations (2.22±1.67 ppm) exceeded FEPA limits, while PM_{2.5} levels (337.28±420.19 µg/m³) surpassed standards set by NESREA, WHO, and USEPA. CO₂, CH₄, NO₂, and SO₂ levels were within permissible limits. Relative humidity negatively influenced PM_{2.5} and most gases ($p < 0.01$), except NH₃. Wind speed had a significant negative effect ($p < 0.10$) on PM_{2.5}, NO₂, NH₃, H₂S, and SO₂, while temperature negatively impacted CH₄, NO₂, and NH₃, but positively affected SO₂ and PM_{2.5}. Health assessments revealed high prevalence of symptoms among workers, including headache (86.8%), tiredness (86.8%), nasal irritation (71.1%), and eye irritation (47.7%). Lung function tests indicated 10% of workers had obstructive patterns (FEV₁/FVC: 86.84±18.32%), while the control group had normal values (98.82±1.52%). PEFR was significantly lower in workers (61.12±27.85%) compared to controls (88.41±21.76%), with 13.3% showing severe airway narrowing. The study concludes that poultry farm air quality is poor and significantly impacts workers' respiratory health, increasing their risk of lung function impairment and airway obstruction due to prolonged pollutant exposure.

KEY WORDS: Poultry production; Air quality; Agriculture; Human health

1. Introduction

One major source of animal protein and a significant contributor to the Nigerian economy is poultry production, which is a subsector of livestock production and accounts for 19% of the country's meat supply (SAGTAP, 2012). The

Nigerian poultry business, according to Sahel (2015), is projected to worth 80 billion (\$600 million) with 165 million birds in it. With over 25 million people employed directly and indirectly in the commercial poultry industry, which has

gradually grown from small backyard farms to large confined structures with intensive management systems. The poultry industry is considered to be the most industrialized segment of the livestock sub-sector (Bello *et al.*, 2015). According to Adene and Oguntade (2006) and Abimiku (2008), the poultry industry in Nigeria is divided into small-scale (500-2,500), medium-scale (2,500-10,000) and large-scale (over 10,000 birds).

The expansion of poultry production resulting to Confined Feeding Animal Operations (CFAOs) is increasingly recognized as a source of air pollutants that have significant environmental and health impacts in and around poultry facilities (Copeland, 2014). This is mainly due to litter and manure generated during production, which pose a serious risk of air pollution arising from the emission of unpleasant odours and microorganisms. According to Akinbile (2012), harmful gas pollutants such as NH₃, CO₂, O₃, N₂O, and other gases are released and contribute between 3–8% to global warming, exacerbating the effects of climate change.

In poultry manure, 57% of the total nitrogen is lost via volatilization within 14 days of dumping (Adeoye *et al.*, 2004). Ammonia volatilization can increase greenhouse gas emissions, generate acid rain, and suffocate people (McGinn and Janzen, 2018). According to Oguntoke *et al.* (2010), indoor air pollution causes a bigger health risk on a worldwide scale than contaminated outdoor air pollution does while a source of air pollution is the unregulated dumping of waste inside and outside the poultry pens. Toxic gases (NH₃, CO₂, and H₂S), odours, dust, and microorganisms are found in poultry housing and are known to have a negative impact on poultry health.

Objectives of the Study

The overall objective was to assess the effect of poultry production on air quality and human health in selected agricultural zones of Imo State.

The specific objectives of this study were to

- i. Assess the gaseous and Particulate Matter pollutants from poultry production systems.
- ii. Determine the microclimatic parameters from intensive poultry production systems and their effect on air pollutants;
- iii. Assess the effects of air pollutants on the health status and lung function of poultry workers.

2. Material and Methods

2.1 Study area

The research area consisted of chicken farms in a few agricultural zones in Imo State, in the southeast of Nigeria. The State was established in February 1976 with 27 Local Government Areas (LGA) and 37 Local Council Development Areas (LCDA) as at the time of writing the reports. Imo State is located between latitude 5°29'0" north and longitude 7°2'0" east. The study area is bordered by Anambra State to the north, Rivers State to the south, Delta State to the west, and Abia State to the east. 4.93 million People live in the state, which has a land area of around 5530 km² (NPC, 2006). The state experiences tropical weather with yearly rainfall ranging from 1600 mm to 900 mm. All year round, the area experiences warm temperatures. Temperatures in the area range from 28°C to 35°C all year round. *Fig. 1* displays the map that depicts the sample at the study area of Imo state.

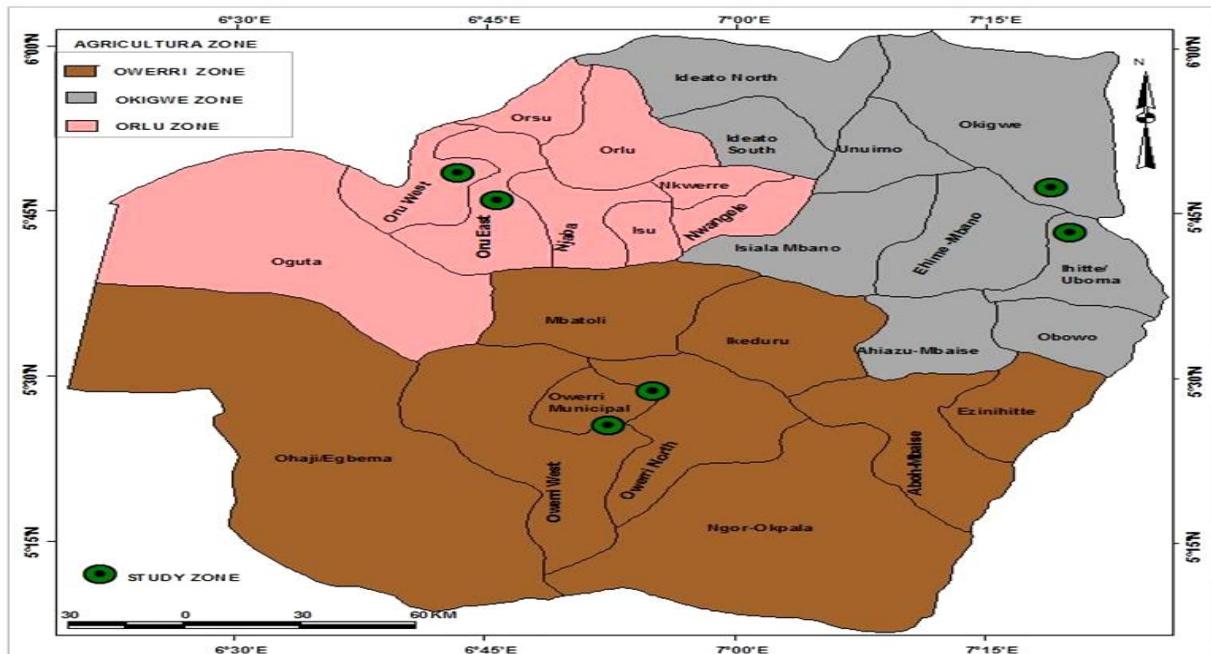


Fig. 1: Map of Imo State, Nigeria, showing the Sampled Locations in the Study Area

2.2 Sample collection and analysis

The Six poultry houses in three zones; Orlu, Okigwe, and Owerri, each marked by the letters R, K, and W as illustrated in Plates 1 - 6 were purposefully chosen from three poultry zones categorized by the Imo State-based Poultry Association of Nigeria (PANIS). Every month from December, 2021 to May, 2022, two poultry houses in each zone were sampled. These poultry houses passed both indoor and outdoor air sampling using the air sampler. In order to achieve a complete coverage of the surroundings for optimal coverage, both inside and outdoor air sampling spots were determined based on the air pollution levels in the poultry houses and population density of the poultry birds. Table 1 displays the parameters of the pens (W_1 , W_2 , K_1 , K_2 , R_1 and R_2) at the time of sampling. The sampled areas outside the pens are designated as W_3 , K_3 , and R_3 .

Thermo PDR 1250 metric sampler was used to detect $PM_{2.5}$ while KanoMax and iTXmultigas Analysers, hand-held air quality monitoring devices, provided direct reading readings of NH_3 , CH_4 , N_2O , H_2S , and CO_2 over the course of an hour at 10-minute intervals. The Kanomax gas analyzer's associated probe was used to acquire meteorological data such as temperature and relative humidity, while the Benetech Model GM816 multipurpose microprocessor digital anemometer was used to monitor wind speed. Monthly sampling was done between December, 2021 and May, 2022.

2.3 Study population

Clearly defined questions to ascertain the extent/level to which air pollution from the production of poultry affects people's health. In addition to asking for information on their

Table 1: Characteristics of Pens across sampled zones in Imo State

Pen	Bird Type	Bird Age (Weeks)	Stock Size	Manure Removal / Frequency	Housing Type	Ventilation Type
W ₁	Layers	45-69	7000	Flushing/ daily	3-tier battery cage	Mechanical and Natural
W ₂	Layers	30-54	20,722	Belt conveyor/ 3 days	4-tier battery cage	Mechanical and Natural
K ₁	Broilers	2-3	9100-10000	Use of shovels/2-3 weeks	Wood shavings litter	Natural
K ₂	Broilers	4-6	5000-10000	3-tier battery cage/4 days	3-tier battery cage	Natural
R ₁	Pullets	15-22	3000	Use of shovel/ 6 weeks	Wood shavings litter	Natural
R ₂	Layers	52-56	1200	Designed to self-flush	3-tier battery cage	Natural

W₁ and W₂ = Sampled Poultry houses in Owerri Zone in Imo State

K₁ and K₂ = Sampled Poultry houses in Okigwe Zone in Imo State

R₁ and R₂ = Sampled Poultry houses in Orlu Zone in Imo State

personal judgment of their health. The purpose of choosing respondents, a purposive sample technique is doubtless. The whole workforce at the three poultry farms comprised the targeted respondents. The demographic distribution between the three poultry farms is shown in [Table 2](#).

Table 2: Population distribution across the three poultry farms

Poultry Zone	Poultry Pen	Number of Workers
Owerri	1	12
	2	14
Okigwe	1	5
	2	5
Orlu	1	1
	2	1
Total		38

2.4 Impacts of air pollutants on the health status of poultry workers

Thirty-eight respondents which represented the total population of workers in the six sampled

poultry pens were administered questionnaires and their responses documented.

2.5 Lung Function Assessment

A hand-held spirometer SP10 in accordance with the ATS standard, respiratory function parameters (FVC, FEV₁, FEV₁/FVC % and PEFR) were assessed (Miller *et al.*, 2020). To help each respondent act appropriately, demonstration exercises of the maneuver were explained to them. Prior to the lung function testing, they were advised to practice this maneuver. Each subject was examined while seated. Each participant's mouthpiece was discarded, and air was forced through it into the spirometer to prevent contamination. The subjects were told to inhale deeply and quickly before exhaling fiercely and thoroughly into the apparatus. The highest FVC, PEFR, and FEV measurements obtained during the first one second were recorded after three readings.



Plate 1: Poultry Pen 1 in Owerri Zone



Plate 2: Poultry Pen 2 in Owerri Zone



Plate 3: Poultry Pen 1 in Okigwe Zone



Plate 4: Poultry Pen 2 in Okigwe Zone



Plate 5: Poultry Pen 1 in Orlu Zone



Plate 6: Poultry Pen 2 in Orlu Zone

2.6 Anthropometric characteristics

Each person from the control group and the poultry workers was questioned about their age in years. A portable stadiometer was used to measure the workers' heights and the weights were determined using a weighing scale.

2.7 Calculation of predicted values of FVC, PEFR and FEV

The observed FVC, PEFR, and FEV values were reported as a percentage of the expected values using regression analysis and a set of prediction equations for adults (Ingle *et al.*, 2005; Reddy *et al.*, 2014; Olujimi *et al.*, 2016).

The following are the prediction equations:
Poultry workers:

$$\begin{aligned} \text{FVC (L)} &= 0.019H - 0.015A - 0.70 \\ \text{FEV1 (L)} &= 0.019H - 0.012A - 1.07 \\ \text{PEFR (L/Sec)} &= -0.008H - 0.121A + 8.45 \end{aligned}$$

Control group:

$$\begin{aligned} \text{FVC (L)} &= 0.037H - 0.014A - 3.82 \\ \text{FEV1 (L)} &= 0.037H - 0.015A - 3.72 \\ \text{PEFR (L/Sec)} &= -0.002H - 0.006A + 6.46 \end{aligned}$$

Where,

H is height in cm

A is age in years.

FVC is Forced Vital Capacity

FEV₁ is Forced Expiratory Volume in one second (1s)

PEFR is Peak Expiratory Flow Rate Litres per minutes (L/min)

Subjects with (FEV₁/FVC) less than 70% were categorised as having an obstructive pattern of lung function defect (Ibhafidon *et al.*, 2014; Lopez *et al.*, 2014).

(ANOVA, t-test, and regression). In order to establish the association between the pollutants and microclimatic factors, the Duncan's Multiple Range Tests and t.test were employed to separate the means of the pollutant concentration variations. Equation (1) depicts the multiple regression model used by to ascertain the association between microclimatic factors and contaminants (Obayelu and Adeniyi, 2006; Nwagwu *et al.*, 2012)

$$Y = a + x_1b_1 + x_2b_2 + x_3b_3 + e \quad \dots \quad (1)$$

Where,

Y = Dependent variables NH₃, CH₄, N₂O, H₂S, CO₂ and PM_{2.5}

x = Independent variables:

x₁ = relative humidity

x₂ = temperature

x₃ = wind speed

a = regression constant

b = regression coefficient

e = error term

2.8 Description of activities at studied poultry sites

This section indicates the concentrations of gaseous pollutants CO₂ across the sampling locations from December, 2021 to May, 2022. W₁, W₂, K₁, K₂, R₁, R₂ represent pens while W₃, K₃ and R₃ denote the sampled points outside the pen. Pen K₁ and K₂ had varying age of birds because broilers were only raised for 6 weeks and were restocked at intervals. K₁, K₂ and K₃ were not sampled in January because there were no birds. Also layers in R₂ were sold after three months of sampling and the pullets were transferred to the battery cage in February, as they had reached point of lay. About half of the birds were later

sold, leaving R_1 with no birds for the rest of the sampling months.

The results BDL = Below Detection Limit

DL: Detection Limit

Control NPPW: Non-Poultry Production Workers

3. Results and Discussion

3.1 Levels of CO₂ pollutants from the studied poultry farms

Carbon dioxide (CO₂) indoor concentrations (Mean±Standard deviation) ranged between 1041.33±32.25 and 1971.25±145.96 mg/m³ with the highest concentration W₂ in the month of March and the lowest in W₃ in January as shown in Fig. 2.

3.2 Mean concentrations of gaseous pollutants and PM_{2.5}

The summary of means of air pollutants are presented in the Table 3. Carbon dioxide (CO₂)

concentrations (Mean±Standard deviation) had a range between 1273.52±221.71 and 1545.65±279.30 mg/m³ with the highest at W₁ and the lowest at W₃ and were significantly higher in the pens.³

Methane (CH₄) concentrations ranged from 0.06±0.40 to 0.81±1.37 ppm. The CH₄ was however not detected W₃, K₁, K₃, R₁, R₂, R₃. The highest mean concentration was significantly highest in W₁ and lowest in K₂. The battery cage system operated in W₁ allows collection of manure in slurry form in the pit which provides anaerobic condition resulting in CH₄ production unlike manure in the solid form in R₂.

Nitrogen dioxide (NO₂) mean concentrations ranged between 0.032 and 0.034 ppm. The highest concentration was recorded in R₁ and the lowest in K₁. NO₂ concentration was significantly highest in R₁.

Ammonia (NH₃) concentration ranged between 0.23±0.43 and 3.04±1.64 ppm. It was BDL in W₃,

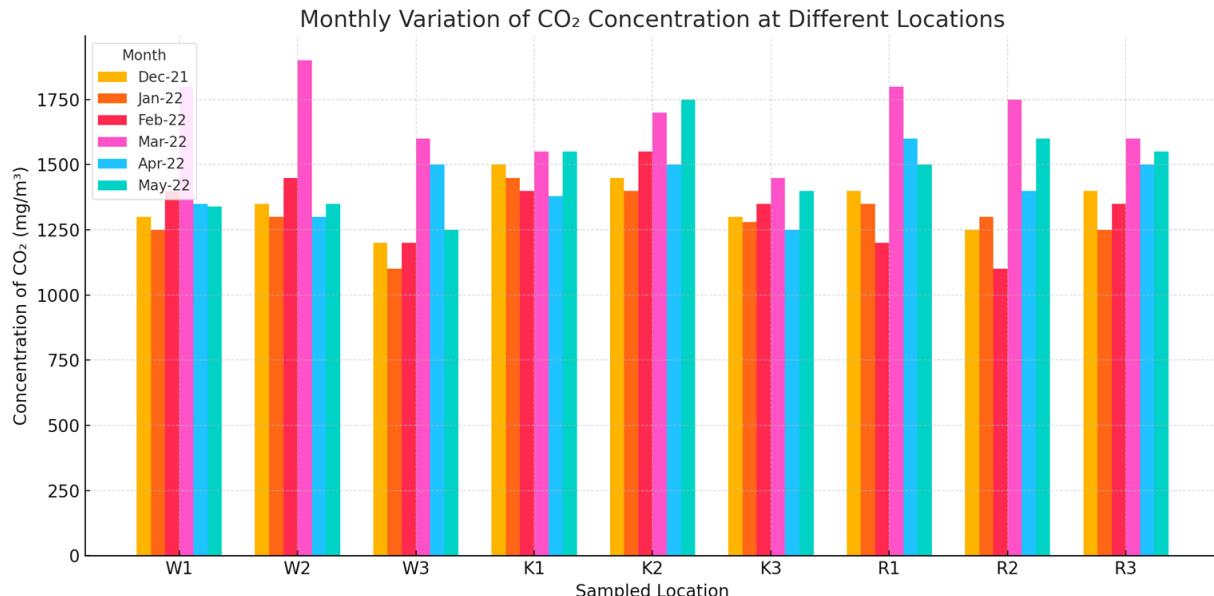


Fig. 2: Monthly variation of CO₂ concentration at different locations

lowest in K₃ and significantly highest in R₂. This may be attributed to the manure management practices. Manure is held back in R₂ for two weeks while the wastes from other battery cage pens are disposed within 2-3 days.

Hydrogen Sulphide (H₂S) concentration ranged from 0.02±0.03 to 0.4±0.88 ppm. H₂S was below detection limit in R₁ and R₃, lowest at R₂ and significantly highest in K₂.

Sulphur Oxide (SO₂) concentration had a range of 0.01±0.01 and 0.44±0.90 ppm. It was BDL in R₁, lowest in K₁ and was significantly higher in zone K₂ and K₃ compared to others. This can be attributed to outdoor source of the emissions from the generator set of the farm which were in operation at the time of sampling.

Particulate Matter (PM_{2.5}) concentrations ranged from 97.90±50.54 to 1469.70±423.44 µg/m³. It was lowest at W₃ but was statistically highest in R₂. This can be attributed to the poultry

Table 3: Mean concentrations of gaseous pollutants and PM_{2.5}

Location	CO ₂ (mg/m ³)	CH ₄ (ppm)	NO ₂ (ppm)	NH ₃ (ppm)	H ₂ S(ppm)	SO ₂ (ppm)	PM2.5 (µg/m ³)
W1	1545.65±279.30 ^a	0.81±1.37 ^a	0.033±0.001 ^b	2.73±1.86 ^{ab}	0.04±0.05 ^c	0.03±0.78 ^b	222.94±228.46 ^c
W2	1466.82±295.91 ^a	0.06±0.40 ^b	0.033±0.001 ^b	2.00±1.5 ^c	0.34±0.83 ^b	0.33±0.06 ^b	222.77±259.26 ^c
W3	1273.52±221.7 ^b	BDL	0.033±0.001 ^b	BDL	0.02±0.02 ^c	0.04±0.05 ^b	97.90±50.54 ^d
K1	1493.94±206.81 ^a	BDL	0.032±0.001 ^c	2.38±1.36 ^b	0.02±0.03 ^c	0.01±0.01 ^b	434.39±399.05 ^b
K2	1476.04±262.18 ^a	0.18±0.45 ^b	0.033±0.001 ^b	2.20±1.48 ^c	0.4±0.88 ^a	0.38±0.83 ^a	269.92±241.61 ^b
K3	1303.97±128.82 ^b	BDL	0.033±0.001 ^b	0.23±0.43 ^e	0.33±0.67 ^b	0.44±0.90 ^a	125.61±91.63 ^d
R1	1353.22±225.0 ^b	BDL	0.034±0.002 ^a	1.36±1.12 ^d	BDL	BDL	1469.70±423.44 ^a
R2	1475.17±264.85 ^a	BDL	0.033±0.001 ^b	3.04±1.64 ^a	0.02±0.03 ^c	0.04±0.9 ^b	321.88±278.52 ^b
R3	1353.19±226.03 ^b	BDL	0.033±0.001 ^b	0.25±0.44 ^e	BDL	0.01±0.03 ^b	179.07±212.98 ^d
Mean	1476.04	0.20	0.033	2.22	0.10	0.10	337.28
SD					0.42	0.42	420.19
Range	262.18	0.72	0.001	1.67	0.02-0.4	0.01-0.44	97.90-1469.70
NESREA	1273.52-1545.65	0.06-0.81	0.032-0.034	0.23-3.04	0.10	0.10	250
WHO(2005)	NA	NA	0.04-0.06	0.3	0.19	0.06	25
USEPA(2001)	900	0.06	0.11	0.5			35

Values are Mean±SD. Values with similar superscripts along the same column are not significantly (p > 0.05) different according to Duncan Multiple Range Test, BDL- Below Detection limit of 0.01ppm/mg/m³, NA- Not Available, SD- Standard deviation

management systems. Pullets of 6 months were raised on litter (wood shavings and dust). This promotes behavioural expression of birds such as scratching and dust bathing. On the other hand, W₁ and W₂ had the lowest PM_{2.5} concentrations across the pens because both combined mechanical and natural ventilation while other pens have only natural ventilation. Hence, increase in air velocity helps to disperse pollutants, and reduce their concentrations as observed in W₁ and W₂.

3.3 Mean values of the microclimatic parameters

Table 4 shows the summary of the mean values of microclimatic parameters of all the poultry sites.

The relative humidity (Mean±SD) ranged from 32.75±8.53 to 59.57±11.44%. The highest and the lowest RH were obtained in W₂ and R₁ respectively. W₁ and W₂ had significantly higher relative humidity than others.

Temperature (Mean±SD) values ranged between 30.74±3.50 and 34.08±2.71 °C. The highest temperature was recorded at K₃ and the lowest at W₂. R₁ and R₂ had significantly higher temperatures than pens.

Wind speed (Mean±SD) ranges from the poultry sites were between 0.05±0.22 and 1.8±0.9 m/s. The highest wind speed was obtained at R₃ while the lowest was at K₂. This may be linked to low/still air movement since temperature was mostly high during sampling period. Pens W₁ and W₂ had significantly higher wind speed which can be attributed to the combination of both mechanical and natural ventilation system adopted compared to other pens.

4. Discussion

The results of the values obtained in the pens are similar to that which was obtained in confined poultry houses in studies conducted by Oriola *et al.* (2013) in Ibadan, Oyo State and Zhao *et al.*

Table 4: Mean values of microclimatic parameters in poultry sites

Sampled Sites	Relative Humidity (%)	Temperature (°C)	Wind Speed (m/s)
W1	58.34±9.12 ^a	31.73±2.79 ^{bcd}	1.20±0.76 ^b
W2	59.57±11.44 ^a	30.74±3.50 ^d	0.83±0.61 ^b
W3	56.58±13.79 ^{ab}	33.06±2.99 ^{ab}	0.22±0.50 ^{cd}
K1	56.88±12.42 ^{ab}	31.13±5.26 ^{cd}	0.06±0.20 ^d
K2	50.88±15.44 ^b	33.53±2.36 ^{ab}	0.05±0.22 ^d
K3	52.39±7.17 ^b	34.25±24.29 ^a	0.70±0.57 ^{bc}
R1	32.75±8.53 ^d	34.08±2.71 ^a	0.48±0.48 ^{cd}
R2	41.57±14.74 ^c	32.38±4.36 ^{bc}	0.15±0.29 ^d
R3	41.89±13.61 ^c	33.94±5.6 ^a	1.81±0.9 ^a
Mean	51.39	31.59	0.54
SD	15.29	4.00	1.54
Range	32.75-58.34	30.74-34.25	0.04-1.8

Note: Values are Mean±SD

Values with similar superscripts along the same column are not significantly (p >0.05) different according to Duncan Multiple Range Test

SD-Standard deviation

(2015). However, NH_3 levels recorded in most pens exceeded the limit of 0.3 ppm NESREA tolerance limit. H_2S was however BDL in all poultry sites in December. These H_2S concentrations were below that which was recorded by Nwagwu *et al.* (2011) in the poultry pens in Port Harcourt, Nigeria during the dry season. The H_2S concentrations were however above the NESREA (2011) tolerant limits of 0.10 ppm in few pens (W_1 , W_2 and K_2), and also exceeded the WHO limits of indoor H_2S (0.06 ppm). Sulphur dioxide (SO_2) concentrations were BDL in the months of December and January in the sampled poultry site. Mean concentrations were however above NESREA (2011), WHO (2015) and USEPA (2011) air quality standard of 0.10 ppm, 0.19 ppm and 0.5 ppm respectively in pens K_2 , K_2 and R_2 . High concentrations as clearly observed at K_2 and K_3 was also reported by Iyogun *et al.* (2018) who attributed the high concentration to combined outdoor and indoor sources of SO_2 . This can be linked to the emissions from the generator set of the farm which was in operation at the time of sampling. Concentrations of $\text{PM}_{2.5}$ in most pens were above the recommended WHO (2015), USEPA (2011) and NESREA (2011) limits of $25 \mu\text{g}/\text{m}^3$, $35 \mu\text{g}/\text{m}^3$ and $250 \mu\text{g}/\text{m}^3$.

The high concentration in January can be associated with combination of dust generated in the poultry houses and harmattan dust. During the sampling in January, 2021, harmattan was quite severe; hence the prevalence of harmattan dust must have had its cumulative effect on the recorded $\text{PM}_{2.5}$ concentrations. This supports the studies conducted on PM pollution in Nigeria during the harmattan period (Efe, 2008; Obioh *et al.*, 2008).

5. Conclusion

The research confirms that poultry production has a significant negative effect on air quality and human health in selected agricultural zones of Imo State, Nigeria. Addressing this issue requires the cooperation and commitment of all stakeholders involved to implement sustainable practices and regulations that prioritize environmental conservation and human well-being.

6. Reference

Abimiku, A. E. (2008). *Policy issues in livestock development in Nigeria*. Jos: JAMB Press.

Adebawale, O. O., & Adeyemo, O. K. (2016). Work characteristics and occupational health hazards of poultry workers in Nigeria. *African Journal of Environmental Science and Technology*, 10(8), 258–266. <https://doi.org/10.5897/AJEST2016.2119>

Adene, D. F., & Oguntade, A. E. (2006). *The structure and importance of the commercial and village-based poultry industry in Nigeria*. Rome: FAO.

Adeoye, G. O., Shridar, M. K. C., & Mohammed, O. E. (2004). Poultry waste management for crop production: Nigerian experience. *Waste Management and Resources*, 22, 165–172.

Adeoye, G. O., Sridhar, M. K. C., Ogunwale, J. A., Makinde, E. A., & Ajayi, F. D. (2004). Evaluation of spent compost manure from mushroom production for crop improvement. *Nigerian Journal of Horticultural Science*, 9, 14–21.

Akinbile, C. O. (2012). Environmental impact of landfill on groundwater quality and agricultural

soils in Nigeria. *Soil and Water Research*, 7(1), 18–26. <https://doi.org/10.17221/8/2011-SWR>

Bello, K. O., Alebiosu, L. A., Lala, A. O., Irekhore, O. T., & Oduguwa, O. O. (2015). Characteristics of commercial poultry and spatial distribution of metabolic and behavioural diseases in Oyo State, Nigeria. *Journal of Veterinary Sciences*, 13(3), 31–41.

Carruth, A. K., Carver, A., & Moffett, B. (2008). Personal protective equipment use among poultry workers in southern U.S. states. *Journal of Occupational and Environmental Hygiene*, 5(9), 611–616.

Copeland, C. (2014). *Air quality issues and animal agriculture: A primer*. Congressional Research Service. <https://sgp.fas.org/crs/misc/RL32948.pdf>

Efe, S. I. (2008). Spatial distribution of particulate air pollution in Nigerian cities: Implications for human health. *Journal of Environmental Health Research*, 7(2), 107–116.

Federal Environmental Protection Agency (NESREA). (2011). *National interim guidelines and standards for industrial effluents, gaseous emissions and hazardous wastes*. Environmental Pollution Control Handbook. NESREA. (pp. 62–67).

Food and Agriculture Organization (FAO). (2008). *Consultative mission on assessment of the Nigerian poultry market chain to improve biosecurity*. <http://www.fao.org/docrep/012/ak778e/ak778e00>

Ibhafidon, L. I., Obaseki, D. O., Erhabor, G. E., Akor, A. A., Irabor, I., & Obioh, I. B. (2014). Respiratory symptoms, lung function and particulate matter pollution in residential indoor environment in Ile-Ife, Nigeria. *Nigerian Medical Journal*, 55(1), 48–53.

Ingle, S. T., Pachpande, B. G., Wagh, N. D., Patel, V. S., & Attarde, S. B. (2005). Exposure to vehicular pollution and respiratory impairment of traffic policemen in Jalgaon City, India. *Indian Health*, 43, 656–662.

Iyogun, C. A., Adewumi, J. R., & Fashola, O. T. (2018). Analysis of sulphur dioxide emissions in poultry facilities in Nigeria. *Environmental Pollution and Control Journal*, 21(4), 77–84.

Kearney, G. D., Shaw, R., Prentice, M., & Tutor-Marcom, R. (2014). Evaluation of respiratory symptoms and respiratory protection behaviour among poultry workers in small farming operations. *Journal of Agromedicine*, 19, 162–170.

Lopez, A. D., Shibuya, K., Rao, C., Mathers, C. D., Hansell, A. L., Held, L. S., Schmid, V., & Buist, S. (2014). Chronic obstructive pulmonary disease: Current burden and future projections. *European Respiratory Journal*, 27(2), 397–412. <https://doi.org/10.1183/09031936.05.00024605>

McGinn, S. M., & Janzen, H. H. (2018). Ammonia emissions from poultry manure. In *Greenhouse gas emissions from livestock and poultry manure* (pp. 75–89). Springer. https://doi.org/10.1007/978-3-319-74437-2_5

Miller, M. R., Lloyd, J., & Bright, P. (2002). Recording flow in the first second of a maximal forced expiratory manoeuvre: Influence of frequency content. *European Respiratory Journal*, 19, 530–533.

Nwagwu, J. E., Okonkwo, I. O., & Ajayi, A. O. (2011). Measurement of hydrogen sulfide concentration in poultry farms in Port Harcourt, Nigeria. *Nigerian Journal of Environmental Sciences and Technology*, 5(1), 22–28.

Nwagwu, J. E., Okonkwo, I. O., & Ajayi, A. O. (2012). Assessment of air pollutants and their correlation with meteorological parameters in urban Nigeria. *Environmental Monitoring and Assessment*, 184(7), 4121–4134. <https://doi.org/10.1007/s10661-011-2247-5>

Obayelu, A. E., & Adeniyi, M. O. (2006). *Statistical analysis of environmental data using multiple regression techniques*. *Journal of Environmental Statistics*, 3(2), 45–54.

Obioh, I. B., Oluwole, A. F., Akeredolu, F. A., & Ogunsola, O. J. (2008). Particulate matter levels during harmattan and implications for health in Nigerian cities. *Atmospheric Environment*, 42(35), 8611–8618.

Ogunteke, O., Afolabi, T. A., & Babatunde, A. I. (2010). Indoor air pollution and health risks among rural dwellers in Odeda area, South-Western Nigeria. *Ethiopian Journal of Environmental Studies and Management*, 3(2), 39–45.

Olujimi, O. O., Akeredolu, F. A., & Oketola, A. A. (2016). Health risks of exposure to emissions from a large-scale poultry farm in southwest Nigeria. *Environmental Monitoring and Assessment*, 188, 121. <https://doi.org/10.1007/s10661-016-5107-7>

Olujimi, O. O., Ana, G. R. E. E., Ogunseye, O. O., & Fabunmi, V. T. (2016). Air quality index from charcoal production sites, carboxyhemoglobin and lung function exposure to hydrogen sulphide in livestock operations. *Journal of Agromedicine*, 20, 225–236.

Oriola, K. O., Alade, A. O., & Adekala, O. S. (2013). Evaluation of air quality in a patterns on particulate matter and polycyclic aromatic hydrocarbon concentrations in Roxbury, Massachusetts. *Journal of Exposure Analysis and Environmental Epidemiology*, 13, 364–371. <https://doi.org/10.1038/sj.jea.7500289>

Radon, K., Danuser, B., Iversen, M., Monso, E., Weber, C., Hartung, J., Donham, K., & Nowak, D. (2001). Respiratory symptoms in European animal farmers. *European Respiratory Journal*, 17(4), 747–754. <https://doi.org/10.1183/09031936.01.17407470>

Rylander, R., & Carvalheiro, M. F. (2006). Airways inflammation among workers in poultry houses. *International Archives of Occupational and Environmental Health*, 79(6), 487–490.

SAGTAP (Sheep and Goats Transformation Action Plan). (2012). *Implementation plan for livestock transformation action plan*. Federal Ministry of Agriculture and Rural Development, Abuja, Nigeria. (p. 20)

Sahel. (2015). *An assessment of the Nigerian poultry sector* (Vol. 11). <https://www.sahelcp.com>

Srivastava, R. K., Sarkar, S., & Beig, G. (2014). Correlation of various gaseous pollutants with meteorological parameter (temperature, relative humidity and rainfall). *Global Journal of Science Frontier Research: Health, Environment and Earth Science*, 14(6), 57–65.

Tzanakis, N., Kallergis, K., Bouros, D., Samiou, M., & Siafakas, N. M. (2001). Short-term effects of environmental tobacco smoke on lung function

of healthy nonsmokers. *American Journal of Respiratory and Critical Care Medicine*, 164(5), 763–768.

United States Environmental Protection Agency (USEPA). (2011). *National ambient air quality standards (NAAQS)*. <https://www.epa.gov/criteria-air-pollutants>

World Health Organization (WHO). (2015). *WHO global air quality guidelines: Particulate matter (PM2.5 and PM10), ozone, nitrogen dioxide, sulfur dioxide and carbon monoxide*. WHO. <https://www.who.int/publications/i/item/9789240034228>

World Health Organization. (2000). *Air quality guidelines for Europe* (2nd ed.). WHO Regional Office for Europe. https://www.euro.who.int/__data/assets/pdf_file/0005/74732/E71922.pdf

Zhao, Y., Shepherd, T. A., Li, H., & Xin, H. (2015). Environmental assessment of three egg production systems - Part II: Ammonia, greenhouse gas, and particulate matter emissions. *Poultry Science*, 94(3), 534–543. <https://doi.org/10.3382/ps/peu076>